

A Highly Stereoselective Synthesis of the AB Disaccharide Unit of Olivomycin A¹

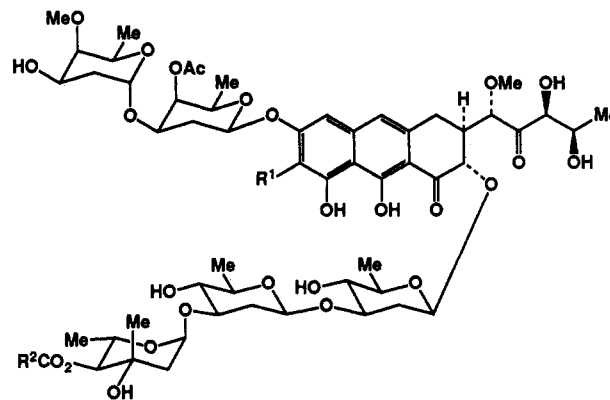
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A highly stereoselective synthesis of the AB disaccharide (4c) of olivomycin A is described. The synthesis features the double asymmetric allylboration of α,β -dialkoxy aldehyde 8 using tartrate allylboronate (*S,S*)-9, which provides triol derivative 10 with the correct relative and absolute stereochemistry for utilization in syntheses of the A and B monosaccharides. The derived diol benzyl ether 11 was converted into 3-hydroxy-2-deoxy-*lyxo*-pyranoside 16 and thioglycoside 18 via the intermediacy of α -methyl 2-deoxy-*lyxo*-pyranoside 14 α , and then 16 and 18 were coupled in CH₂Cl₂ at 23 °C by using Nicolaou's NBS thioglycosidation procedure to give the α,α -disaccharide 4a with >6:1 selectivity. Difficulties encountered in the hydrolysis or activation of the methyl glycoside function of 4a prompted the synthesis to be revised by using intermediates containing more easily deprotected anomeric silyl (TBDMS) ethers. Diol 11 was thus selectively monosilylated to give 21, which was smoothly converted via 2-deoxy-*lyxo*-pyranose 23 into thioglycoside 18 and 2-deoxypyranoside 27, which possesses a TBDMS protecting group at the anomeric center. Intermediates 18 and 27 were coupled again by using the Nicolaou methodology in a CH₃CN–THF cosolvent mixture at –42 °C to give the AB disaccharide TBDMS acetal 4b with $\geq 20:1$ selectivity. Deprotection of the anomeric TBDMS acetal by using Et₃NH⁺F⁻ in CH₃CN then completed the synthesis of the olivomycin AB disaccharide unit 4c.

Olivomycin A (1), chromomycin A₃ (2), and mithramycin (3) are clinically active members of the aureolic acid family of anticancer agents.² These compounds are inhibitors of DNA-dependent RNA polymerase, and available structure–activity data indicate that the two oligosaccharide chains are essential for biological activity, influencing both the DNA binding profile as well as cellular uptake.^{3,4} These factors, among others, have stimulated considerable interest in the total synthesis of these structurally complex antibiotics, the ultimate goal being the development of potentially less toxic analogues.⁵ Several syntheses of the olivomycin aglycon, olivin, or its tri-*O*-methyl derivative, have been reported,⁶ and Thiem has published a substantial body of work on the synthesis of the oligosaccharide fragments of 1–3, which also serve as a rigorous proof of the oligosaccharide stereostructures.^{7,8} Other



1, R¹ = H, R² = *i*-Pr, olivomycin A
2, R¹ = R² = Me, chromomycin A₃

(1) Portions of this research were performed at the Massachusetts Institute of Technology, Cambridge, MA, and are described in the Ph.D. Thesis of J. A. Straub (M.I.T., 1987). We gratefully acknowledge a grant from the National Cancer Institute (CA 29847) for support of the initial phase of these studies.

(2) (a) Remers, W. A. *The Chemistry of Antitumor Antibiotics*; Wiley-Interscience, New York, 1979; Chapter 3. (b) Skarbek, J. D.; Speedie, M. K. In *Antitumor Compounds of Natural Origin: Chemistry and Biochemistry*; Aszalos, A., Ed; CRC Press: Boca Raton, FL, 1981; Chapter 5. (c) Pettit, G. R. *Biosynthetic Products for Cancer Chemotherapy*; Plenum Press, New York, 1977; Vol. 1, p 143.

(3) Chromomycin is reported to bind in the DNA minor groove as a 2:1 dimer: (a) Gao, Z.; Patel, D. *Biochemistry* 1989, 28, 751. (b) Banville, D. L.; Keniry, M. A.; Kam, M.; Shafer, R. H. *Ibid.* 1990, 29, 6521.

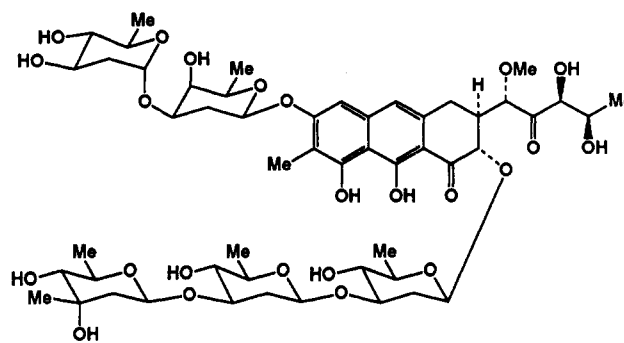
(4) Gupta, R. S. *J. Cell. Physiol.* 1982, 113, 11.

(5) For a review of synthetic efforts through 1987, see: Franck, R. W.; Weinreb, S. M. In *Studies in Natural Product Chemistry*; Rahman, A. U., Ed.; Elsevier: Amsterdam, 1989; pp 173–208.

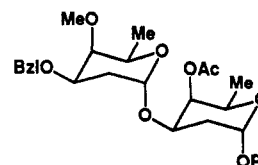
(6) (a) Racemic tri-*O*-methylolivin: Dodd, J. H.; Starrett, J. E., Jr.; Weinreb, S. M. *J. Am. Chem. Soc.* 1984, 106, 1811. (b) Optically active tri-*O*-methylolivin: Franck, R. W.; Bhat, V.; Subramaniam, C. S. *Ibid.* 1986, 108, 2455. (c) Olivin in the naturally occurring enantiomeric form: Roush, W. R.; Michaelides, M. R.; Tai, D. F.; Chong, W. K. M. *Ibid.* 1987, 109, 7575. (d) Roush, W. R.; Michaelides, M. R.; Tai, D. F.; Lesur, B. M.; Chong, W. K. M.; Harris, D. J. *Ibid.* 1989, 111, 2984.

(7) Oligosaccharide stereostructural assignments: (a) Thiem, J.; Meyer, B. *J. Chem. Soc., Perkin Trans.* 1979, 1331. (b) Thiem, J.; Meyer, B. *Tetrahedron* 1981, 37, 551. (c) Thiem, J.; Schneider, G. *Angew. Chem., Int. Ed. Engl.* 1983, 22, 58.

(8) Recent synthetic studies: (a) Thiem, J.; Gerken, M. *J. Org. Chem.* 1985, 50, 954. (b) Thiem, J.; Schoettmer, B. *Angew. Chem., Int. Ed. Engl.* 1987, 26, 555. (c) For a review, see: Thiem, J. In *Trends in Synthetic Carbohydrate Chemistry*; Horton, D.; Hawkins, L. D., McGarvey, G. J., Eds; ACS Symposium Series 386, 1989; Chapter 8.

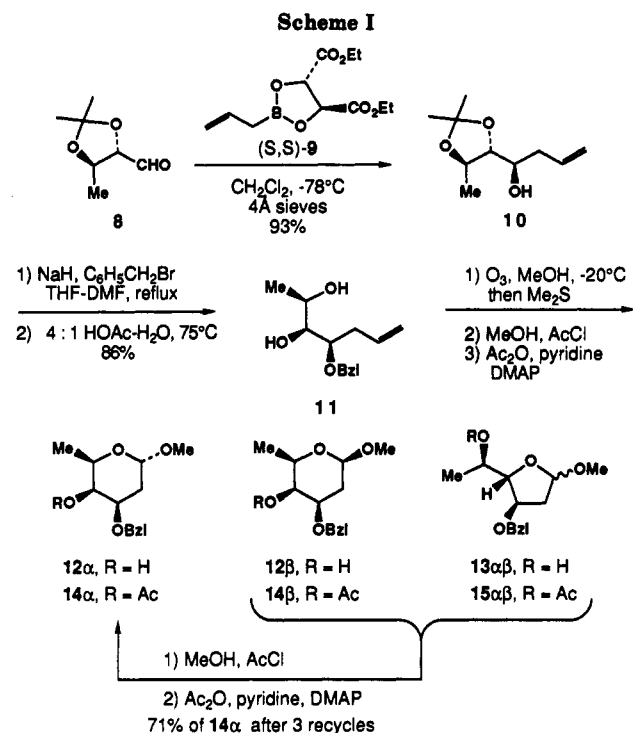


3, mithramycin (aureolic acid)



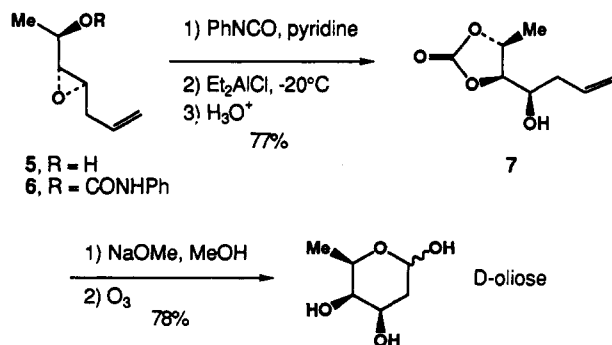
4a, R = Me
4b, R = TBDMS
4c, R = H

studies on the synthesis of the oligosaccharide units have also been reported.⁹



In continuation of our efforts to complete a total synthesis of 1, we have developed an efficient and highly stereoselective synthesis of the AB disaccharide unit (4) of olivomycin A, the details of which are provided herein.¹⁰

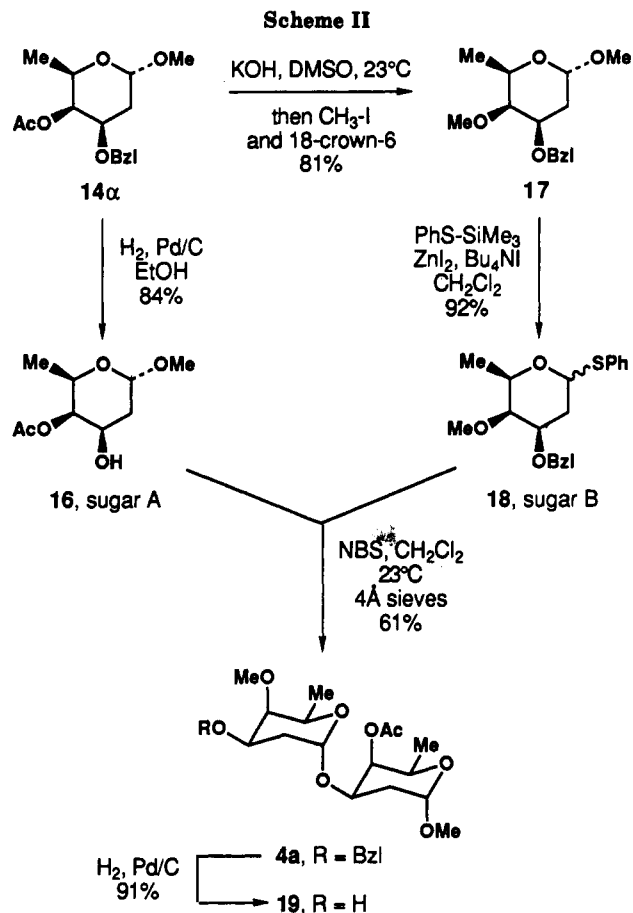
Recognizing that disaccharide 4 consists of two differentially functionalized 2,6-dideoxy-D-lyxo-hexose ("D-oliose") units, we adopted a synthetic strategy that would permit both monosaccharides to be synthesized from a common precursor. Although we had already synthesized D-oliose precursor 7 by the Lewis acid mediated neighboring group promoted reaction of epoxy urethane 6, this route was deemed unsuitable for the present purposes since the synthesis of the key intermediate 5 involves a Sharpless kinetic resolution-asymmetric epoxidation, and the yield of 5 was only 40% on the basis of the racemic allylic alcohol precursor.¹¹ We elected instead to utilize the asymmetric allylboration reaction to establish the stereochemistry of the carbohydrate backbone, anticipating that the synthetic efficiency would be much better if a resolution were avoided.¹²⁻¹⁴



(9) For leading references to other studies on the synthesis of the aureolic acid oligosaccharides, see: (a) Ramesh, S.; Franck, R. W. *J. Chem. Soc., Chem. Commun.* 1989, 960. (b) Binkley, R. W.; Koholic, D. *J. Org. Chem.* 1989, 54, 3577. (c) Ramesh, S.; Kaila, N.; Grewal, G.; Franck, R. W. *Ibid.* 1990, 55, 5 and literature cited therein. (d) Binkley, R. W. *J. Carbohydr. Chem.* 1990, 9, 507.

(10) A preliminary account of this work has appeared: Roush, W. R.; Straub, J. A. *Tetrahedron Lett.* 1986, 27, 3349.

(11) (a) Roush, W. R.; Brown, R. J. *J. Org. Chem.* 1983, 48, 5093. (b) Roush, W. R.; Brown, R. J.; DiMare, M. *Ibid.* 1983, 48, 5083.



The synthesis of disaccharide 4a, the initial target, began with the matched double asymmetric reaction of the readily available chiral aldehyde 8¹⁵ and diethyl tartrate allylboronate (S,S)-9.¹³ This reaction provided the known homoallylic alcohol 10,¹⁶ typically with $\geq 98\%$ stereoselectivity (Scheme I). Benzoylation of 10 followed by hydrolysis of the acetonide gave diol 11 in 86% yield. Ozonolysis of 11 then provided 2,6-dideoxy-D-lyxo-hexose benzyl ether as a mixture of pyranose and furanose tautomers (72% yield). This mixture was converted to a mixture of methyl glycosides 12 α,β and 13 α,β (unseparable) that was directly acylated. Fortunately, α -pyranoside 14 α could be separated at this stage chromatographically (36% yield). The mixture of the β -pyranoside 14 β and the two furanose tautomers 15 α,β , obtained in 48% yield, was re-subjected to the glycosidation conditions to regenerate a mixture of 12 α,β , and 13 α,β that was acylated and again

(12) For reviews of the reactions of allylmetal compounds and aldehydes, see: (a) Hoffmann, R. W. *Angew. Chem., Int. Ed. Engl.* 1982, 21, 555. (b) Roush, W. R. In *Comprehensive Organic Synthesis*; Heathcock, C. H., Ed.; Pergamon Press: Oxford, Vol. 2, in press.

(13) (a) Roush, W. R.; Walts, A. E.; Hoong, L. K. *J. Am. Chem. Soc.* 1985, 107, 8186. (b) Roush, W. R.; Halterman, R. L. *Ibid.* 1986, 108, 294. (c) For fully optimized experimental conditions, see: Roush, W. R.; Hoong, L. K.; Palmer, M. A. J.; Park, J. D. *J. Org. Chem.* 1990, 55, 4109. (d) For a detailed discussion of the stereoselectivity of the reactions of the tartrate allylboronates and alkoxy-substituted aldehydes, see: Roush, W. R.; Hoong, L. K.; Palmer, M. A. J.; Straub, J. A.; Palkowitz, A. D. *Ibid.* 1990, 55, 4117. (e) For the reactions of the chiral crotylboronates and α -methyl chiral aldehydes, see: Roush, W. R.; Palkowitz, A. D.; Ando, K. *J. Am. Chem. Soc.* 1990, 112, 6348.

(14) For a stereochemically general synthesis of 2-deoxyhexoses based on the asymmetric allylboration of 2,3-epoxy aldehydes, see: Roush, W. R.; Straub, J. A.; VanNieuwenhze, M. S. *J. Org. Chem.*, previous paper in this issue.

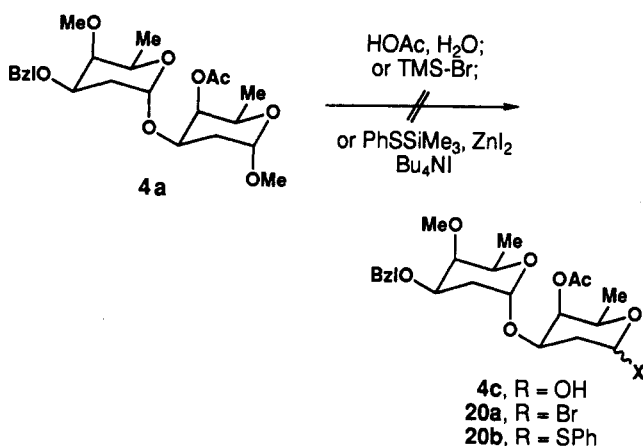
(15) (a) Servi, S. *J. Org. Chem.* 1985, 50, 5865. (b) See also Roush, W. R.; Harris, D. J.; Lesur, B. M. *Tetrahedron Lett.* 1983, 24, 2227.

(16) Fronza, G.; Fuganti, C.; Grasselli, P.; Pedrocchi-Fantoni, G.; Zirotti, C. *Tetrahedron Lett.* 1982, 23, 4143.

separated chromatographically. Recycling of the resulting 14β - $15\alpha,\beta$ mixture two additional times brought the overall yield of 14α to 71%.

Hydrogenation of 14α over 10% Pd on C provided the A-ring monosaccharide **16** in 84% yield. Treatment of 14α with powdered KOH in Me_2SO followed by addition of MeI and 18-crown-6 provided methyl ether **17** in 81% yield (Scheme II), which was then converted into the thioglycoside **18**, an activated form of the B ring monosaccharide, in 92% yield by using Hanessian's procedure.¹⁷ These intermediates were coupled by treatment of a mixture of **17** (1.1 equiv) and **18** (1.0 equiv) with NBS in CH_2Cl_2 at 23 °C in the presence of 4-Å molecular sieves.¹⁸ Although a mixture of anomeric glycosides was anticipated at the outset,^{18,19} we were delighted to find that this procedure provided α,α -disaccharide **4a** in 61% yield with $\geq 6:1$ stereoselectivity. The stereochemistry of **4a** was verified by hydrogenolysis of the benzyl ether that provided the known disaccharide **19**.²⁰

Although this synthesis of **4a** was highly stereoselective and was relatively efficient (10 steps, 17% yield from **8**), the recycle of 14β and $15\alpha,\beta$ notwithstanding, the problem of deprotection and activation of **4a** to provide a suitably activated disaccharide derivative for use in glycosidation reactions with the aglycon olivin remained to be solved. Unfortunately, attempts to hydrolyze **4a** to the free acetal **4c** by using aqueous HOAc,²¹ or attempts to convert **4a** directly to the bromo or thiophenyl glycoside derivatives **20a,b** by using Me_3SiBr ^{20,22} or the Hanessian thioglycosidation protocol,¹⁷ lead to competitive cleavage of the internal α -disaccharide linkage. It thus was clear that a different acetal protecting group was needed for the A-ring monosaccharide unit.



Since the main problem in the synthesis of 14α is the poor kinetic and thermodynamic selectivity for formation

(17) Hanessian, S.; Guindon, Y. *Carbohydr. Res.* 1980, 86, C3.

(18) Nicolaou, K. C.; Seitz, S. P.; Papahatjis, D. P. *J. Am. Chem. Soc.* 1983, 105, 2430.

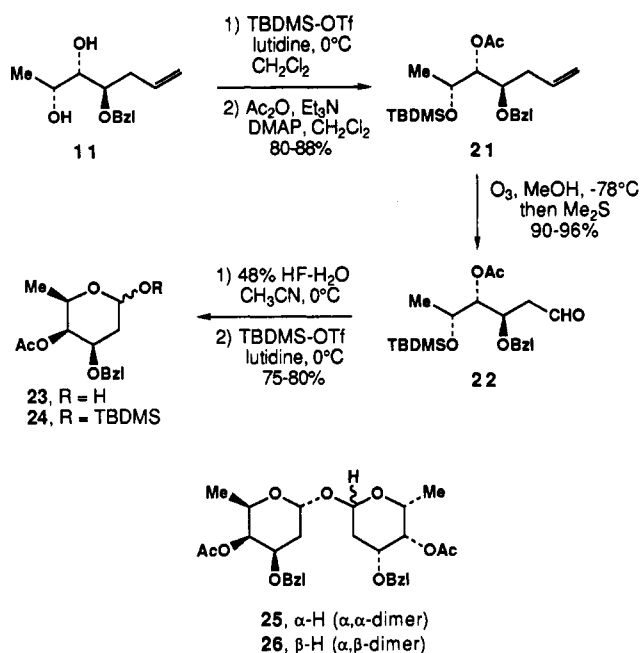
(19) For leading references to other thioglycosidation methods, see: (a) Veeneman, G. H.; van Boom, J. H. *Tetrahedron Lett.* 1990, 31, 275. (b) Dasgupta, F.; Garegg, P. J. *Carbohydr. Res.* 1988, 177, C13. Fügedi, P.; Garegg, P. J. *Ibid.* 1986, 149, C9. (c) Ito, Y.; Ogawa, T. *Tetrahedron Lett.* 1988, 29, 1061. (d) Pozsgay, V.; Jennings, H. J. *J. Org. Chem.* 1987, 52, 4635. (e) Wuts, P. G. M.; Bigelow, S. S. *Ibid.* 1983, 48, 3489. (f) Woodward, R. B.; et al. *J. Am. Chem. Soc.* 1981, 103, 3215. (g) Hanessian, S.; Bacquet, C.; Lehong, N. *Carbohydr. Res.* 1980, 80, C17. (h) Ferrier, R. J.; Hay, R. W.; Vethaviasar, N. *Ibid.* 1973, 27, 55.

(20) Thiem, J.; Meyer, B. *Chem. Ber.* 1980, 113, 3058.

(21) 2-Deoxy- α -glycosides are known to be labile to even to aqueous HOAc under mild conditions. See, for example: (a) Honda, S.; Kakehi, K.; Takai, H.; Takiura, K. *Carbohydr. Res.* 1973, 29, 477. (b) Miyamoto, M.; Kawamatsu, Y.; Shinohara, M.; Nakadaira, Y.; Nakanishi, K. *Tetrahedron* 1966, 22, 2785.

(22) Gillard, J. W.; Israel, M. *Tetrahedron Lett.* 1981, 22, 513.

Scheme III

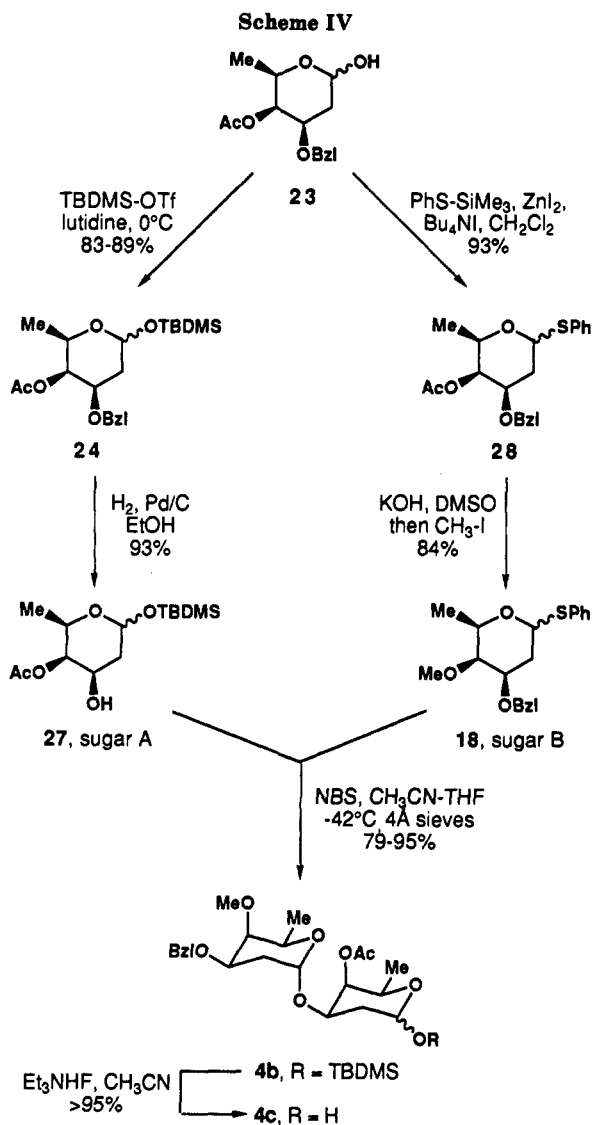


of the desired α -pyranoside from the dihydroxyaldehyde generated by ozonolysis of **11**,²³ we decided to differentiate the C(5) and C(6) hydroxyl groups of **11** prior to this step. Selective silylation of **11** was smoothly accomplished by using *tert*-butyldimethylsilyl triflate (TBDMS-OTf) and lutidine in CH_2Cl_2 , providing the C(6)-monosilyl ether derivative in 82–90% yield along with a small amount (ca. 4%) of disilylated material (Scheme III). Acylation of the C(6)-monosilyl ether then provided **21** in 80–88% overall yield. Ozonolysis of **21** produced the fully protected aldehyde **22** in 90–96% yield that when treated with 48% aqueous HF in CH_3CN at 0 °C cyclized to the targeted hexose **23** (85–90% yield). Also obtained were small amounts of the structurally interesting α,α - and α,β -dimers **25** and **26**. It was not possible to suppress the formation of **25** and **26** by removing the TBDMS protecting group prior to the ozonolysis of **21** owing to the intervention of an extremely facile acyl migration reaction that produced a mixture of regioisomeric acetates. Silylation of **23** using TBDMS-OTf then provided **22** (as an anomeric mixture) in excellent yield.

Recognizing that **22** and **24** are structural isomers, several attempts were made to convert **22** directly into **24** without the stoichiometric removal and reintroduction of the TBDMS group. Treatment of **22** with catalytic amounts of various Lewis acids including TBDMS-OTf or TBDMS-Cl in CH_3CN or THF at temperatures between -78 and -20 °C indeed accomplished the desired conversion. Unfortunately, however, significant amounts of dimers **25** and **26** were also produced. Consequently, the most efficient means of preparing **24** involves the two-step conversion proceeding via **23**.

With convenient and efficient syntheses of **23** and **24** in hand, we turned to the completion of the synthesis of the AB disaccharide **4c**. Thus, hydrogenation of **24** provided the desired A-ring monosaccharide **27** in 93% yield (Scheme IV). Thioglycoside **28** was prepared in 93% yield from **23** by using Hanessian's procedure.¹⁷ Sequential treatment of **28** with KOH in DMSO to hydrolyze the

(23) This problem is well known for sugars in the galactose series (as is **14a**): Pater, R. H.; Coelho, R. A.; Mowery, D. F., Jr. *J. Org. Chem.* 1973, 38, 3272.



acetyl group followed by addition of excess MeI then provided thiosugar 18 in 84% yield. The coupling of 27 and 18 was performed by using Nicolaou's NBS protocol as described for the synthesis of 4a,¹⁸ with the exception that it was performed in a 2:1 mixture of CH₃CN and THF at -42 °C. This reaction proved to be very fast and extremely selective; disaccharide 4b was obtained in excellent yield (79–95%) with very high selectivity for the desired α -glycosidic linkage. The only instance in which a product with a β -glycosidic linkage was detected was one experiment in which the purified β -anomer of 27 was used as the glycosyl acceptor: this experiment provided 79% of 4b along with 4% of a minor product that was identified as the β , β -isomer. Finally, the TBDMS group of 4b was easily removed by treatment with Et₃NH⁺F⁻ in CH₃CN,²⁴ thus providing the targeted AB disaccharide unit 4c in essentially quantitative yield.

In summary, we have developed a highly stereoselective synthesis of 4c that corresponds to the AB disaccharide units of olivomycin A (1) as well as chromomycin A₃ (2). Efforts to complete a total synthesis of 1 are ongoing and additional progress will be reported in due course.

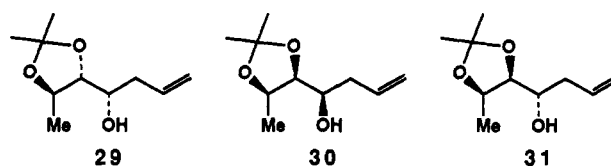
Experimental Section

General. All reactions were conducted in oven-dried (125 °C)

glassware under atmospheres of dry argon or nitrogen. All solvents were purified before use. Ether, tetrahydrofuran (THF), and toluene were distilled from sodium benzophenone ketyl; CH₂Cl₂ and acetonitrile were distilled from CaH₂, methanol from Mg turnings. Removal of solvents was performed by using a rotary evaporator at reduced pressure.

Analytical thin layer chromatography (TLC) was performed by using Kieselgel 60 F₂₅₄ glass plates precoated with a 0.25-mm thickness of silica gel. Column chromatography was performed on kieselgel 60 (230–400 mesh) silica gel according to the method of Still.²⁵ Preparative TLC was performed on kieselgel 60 F₂₅₄ glass plates precoated with a 0.50-mm thickness of silica gel. Unless noted otherwise, all compounds isolated by chromatography were sufficiently pure (>95% according to ¹H NMR analysis) for use directly in subsequent preparative reactions.

lyxo-Hept-1-ene-4,5,6-triol 5,6-Acetonide (10). A solution of aldehyde 8 (1.25 g, 8.70 mmol)¹⁵ in CH₂Cl₂ (7 mL) was added dropwise to a -78 °C solution of (*S,S*)-diethyl tartrate allylboronate 9 (2.88 g, 11.3 mmol)^{13b,14} and 4-Å molecular sieves (200 mg) in CH₂Cl₂ (15 mL). The reaction mixture was stirred at -78 °C overnight and then was allowed to warm to 23 °C. Aqueous NaOH (1 N, 10 mL) was added, and the mixture was stirred for 45 min. The aqueous phase was extracted with ether (3 × 20 mL). The combined organic extracts were dried (MgSO₄), filtered, and concentrated in vacuo. Analysis of the crude product by GC (10-ft Chromosorb G on Carbowax column; 80°/2 min, then 10°/min ramp to 190 °C) indicated that the ratio of 10 (*t*_R 11.5 min) and its hydroxyl epimer 29 (*t*_R 10.5 min) was 98:2. Also present were



small amounts (1–5% total, variable from run to run) of epimers 30 (*t*_R 10.9 min) and 31 (*t*_R 11.9 min) that derive from the C-(2)-epimer of 8. The crude product was chromatographed (20% EtOAc/hexanes) to give 1.51 g (93%) of 10 as a colorless liquid; *R*_f 0.32 (20% EtOAc/hexanes); [α]_D²² -2.6° (*c* = 0.89, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 5.87–5.78 (m, 1 H), 5.18–5.12 (m, 2 H), 4.09 (dq, *J* = 7.8, 6.1 Hz, 1 H), 3.72 (tdd, *J* = 5.5, 5.5, 3.8 Hz, 1 H), 3.48 (dd, *J* = 7.8, 5.5 Hz, 1 H), 2.39–2.34 (m, 1 H), 2.22–2.15 (m, 1 H), 2.18 (d, *J* = 3.8 Hz, 1 H, OH), 1.38 (s, 3 H) 1.36 (s, 3 H), 1.32 (d, *J* = 6.1 Hz, 3 H); IR (neat) 3460, 3060, 1645 cm⁻¹; MS *m/z* 186 (M⁺). Anal. Calcd for C₁₀H₁₈O₃: C, 64.49; H, 9.74. Found: C, 64.20; H, 9.81.

lyxo-4-O-Benzylhept-1-ene-4,5,6-triol (11). A solution of alcohol 10 (7.87 g, 42.2 mmol) in dry THF (50 mL) was added dropwise via syringe over 10 min to a 0 °C suspension of NaH (2.33 g, 57% in oil, 55.3 mmol) in THF (50 mL). The cooling bath was removed and the mixture stirred at 23 °C for 1 h. The mixture was then cooled to 0 °C and neat benzyl bromide (15.3 mL, 129 mmol) was added. The mixture was then stirred at 23 °C for 24 h. The reaction was quenched by the slow addition of water (ca. 20 mL) and diluted with ether (100 mL). The organic layer was washed with water (2 × 100 mL) and brine (2 × 100 mL). The combined aqueous layers were extracted with ether (2 × 150 mL). The combined organic extracts were dried over MgSO₄ and then filtered and concentrated in vacuo. The crude product was purified by flash column chromatography (hexane to elute excess benzyl bromide, followed by 6% EtOAc–hexane to elute the product), giving 0.3 g of recovered 10 (4%) and 10.2 g (88%) of the product benzyl ether as a pale yellow liquid; *R*_f 0.48 (10% EtOAc/hexane); [α]_D²² -33.1° (*c* = 0.51, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.39–7.27 (m, 5 H), 5.97–5.87 (m, 1 H), 5.18–5.09 (m, 2 H), 4.66 and 4.58 (AB dd, *J* = 11.4 Hz, 2 H), 4.06 (dq, *J* = 7.6, 6.1 Hz, 1 H), 3.63 (dd, *J* = 7.6, 6.1 Hz, 1 H), 3.58 (dt, *J* = 6.1, 4.5 Hz, 1 H), 2.50–2.38 (m, 2 H), 1.43 (s, 3 H), 1.40 (s, 3 H), 1.33 (d, *J* = 6.1 Hz, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 138.2, 134.3, 128.3, 127.8, 127.6, 108.0, 82.9, 79.2, 75.0, 72.3, 35.6, 27.3, 26.9, 19.4; IR (neat) 2970, 1645, 1605, 1450, 1375 cm⁻¹; MS *m/z*

(24) Hünig, S.; Wehner, G. *Synthesis* 1975, 180.

(25) Still, W. C.; Kahn, M.; Mitra, A. *J. Org. Chem.* 1978, 43, 2923.

261 ($M^+ - CH_3$). Anal. Calcd for $C_{17}H_{24}O_3$: C, 73.88; H, 8.75. Found: C, 73.51; H, 8.51.

A solution of the above benzyl ether (2.08 g, 7.52 mmol) in 80% aqueous HOAc (15 mL) was heated to 75 °C for 4 h. Most of the solvent was removed in vacuo and the residual HOAc was removed by coevaporation with heptane. The residue was purified by flash column chromatography (45% EtOAc/hexanes) to give 98 mg of recovered benzyl ether (5%) and 1.55 g of 11 (87% yield) as a pale yellow liquid: R_f 0.34 (45% EtOAc/hexanes); $[\alpha]_D^{25} -19.8^\circ$ ($c = 0.33$, $CHCl_3$); 1H NMR (500 MHz, $CDCl_3$) δ 7.40–7.27 (m, 5 H), 5.92–5.82 (m, 1 H), 5.21–5.08 (m, 2 H), 4.68 and 4.54 (AB dd, $J = 11.3$ Hz, 2 H), 4.08 (qdd, $J = 6.4, 2.9, 2.4$ Hz, 1 H), 3.68 (td, $J = 6.1, 4.4$ Hz, 1 H), 3.41–3.37 (m, 1 H), 2.66 (d, $J = 2.9$ Hz, 1 H for OH), 2.50 (d, $J = 7.9$ Hz, 1 H for OH), 2.52–2.46 (m, 1 H), 2.39–2.33 (m, 1 H), 1.20 (d, $J = 6.4$ Hz, 3 H); IR (neat) 3550, 3000, 2940, 1635, 1480, 1450, 1200, 1080 cm^{-1} ; MS (m/z 236 (M^+)). Anal. Calcd for $C_{14}H_{20}O_3$: C, 71.16; H, 8.79. Found: C, 71.03; H, 8.79.

lyxo-5-Acetoxy-4-(benzyloxy)-6-(tert-butyltrimethylsilyloxy)hept-1-ene (21). To a 0 °C solution of diol 11 (1.55 g, 6.6 mmol) in CH_2Cl_2 (20 mL) were added 2,4-lutidine (2.3 mL, 19.9 mmol) and TBDMS-OTf (1.57 mL, 6.7 mmol) dropwise via syringe. The mixture was stirred at 0 °C for 30 min and then was quenched with brine (50 mL) and diluted with ether (100 mL). The aqueous layer was extracted with 100 mL of ether. The combined organic layers were washed with brine (2 × 50 mL), dried ($MgSO_4$), filtered, and concentrated in vacuo. The crude product was purified by flash chromatography (7% EtOAc/hexanes) to give 112 mg of the disilylated product (3.6%; R_f 0.8) and 2.03 g (88%) of the desired C(6)-mono-TBDMS ether as a colorless liquid: $R_f = 0.50$ (10% EtOAc/hexanes); $[\alpha]_D^{25} -47.0^\circ$ ($c = 1.02$, $CHCl_3$); 1H NMR (500 MHz, $CDCl_3$) δ 7.38–7.27 (m, 5 H), 5.99–5.92 (m, 1 H), 5.20–5.10 (m, 2 H), 4.67 and 4.42 (AB dd, $J = 11.1$ Hz, 2 H), 4.18 (qd, $J = 6.3, 1.9$ Hz, 1 H), 3.46–3.41 (m, 1 H), 3.29 (ddd, $J = 9.0, 8.3, 1.9$ Hz, 1 H), 2.66–2.58 (m, 1 H), 2.54–2.48 (m, 1 H), 2.42 (d, $J = 9.0$ Hz, 1 H for OH), 1.22 (d, $J = 6.3$ Hz, 3 H), 0.90 (s, 9 H), 0.10 (s, 3 H), 0.071 (s, 3 H); IR (neat) 3540, 1635 cm^{-1} ; MS (CI) m/z 351 ($M^+ + 1$).

A solution of the above mono-TBDMS ether (1.99 g, 5.67 mmol) in CH_2Cl_2 (20 mL) at 0 °C was treated with Et_3N (7.9 mL, 56.7 mmol), Ac_2O (2.68 mL, 28.4 mmol), and a catalytic amount of DMAP. The mixture was stirred at 23 °C overnight and then diluted with Et_2O (200 mL) and brine (100 mL). The aqueous layer was separated and extracted with Et_2O (2 × 150 mL). The combined organic extracts were washed with brine (100 mL) and saturated $CuSO_4$ solution (2 × 150 mL) and dried over $MgSO_4$. The filtrate was concentrated in vacuo and the residue purified by chromatography on silica gel (8% EtOAc/hexane), giving 2.22 g of 21 (88% from 11) as a clear colorless liquid: R_f 0.50 (10% EtOAc/hexane); $[\alpha]_D^{25} +36.7^\circ$ ($c = 0.95$, $CHCl_3$); 1H NMR (500 MHz, $CDCl_3$) δ 7.34–7.25 (m, 5 H), 5.91–5.82 (m, 1 H), 5.12–5.07 (m, 2 H), 5.09 (dd, $J = 5.9, 4.3$ Hz, 1 H), 4.64 and 4.49 (AB dd, $J = 11.2$ Hz, 2 H), 4.02 (qd, $J = 6.3, 5.9$ Hz, 1 H), 3.63 (ddd, $J = 6.5, 4.3, 2.6$ Hz, 1 H), 2.41–2.34 (m, 1 H), 2.33–2.27 (m, 1 H), 2.09 (s, 3 H), 1.13 (d, $J = 6.3$ Hz, 3 H), 0.89 (s, 9 H), 0.071 (s, 3 H), 0.055 (s, 3 H); ^{13}C NMR (125 MHz, $CDCl_3$) δ 170.3, 138.1, 135.0, 128.3, 127.9, 127.6, 116.8, 77.2, 76.5, 71.3, 67.3, 34.4, 25.7, 21.1, 20.4, 17.9, -4.3, -4.7; IR (neat) 2960, 1745, 1645, 1235, 1080 cm^{-1} ; MS (CI) m/z 393 ($M^+ + 1$). Anal. Calcd for $C_{22}H_{36}O_4Si$: C, 67.30; H, 9.24. Found: C, 67.29; H, 9.19.

4-O-Acetyl-3-O-benzyl-2,6-dideoxy-D-lyxo-pyranose (23). A -78 °C solution of 21 (1.20 g, 3.05 mmol) in MeOH (20 mL) was treated with a stream of O_3 in O_2 for 8 min. The reaction was carefully monitored by TLC analysis. Excess O_3 was removed by continued flashing with N_2 , and then excess Me_2S (3 mL) was added. This mixture was stirred overnight at 23 °C and then concentrated in vacuo. Purification of the crude product by flash column chromatography on silica gel (15% EtOAc/hexanes) gave 1.16 g of aldehyde 22 (96%) as a colorless viscous oil: R_f 0.40 (20% EtOAc/hexane); $[\alpha]_D^{25} +53.4^\circ$ ($c = 1.9$, $CHCl_3$); 1H NMR (500 MHz, $CDCl_3$) δ 9.74 (dd, $J = 2.4, 1.6$ Hz, 1 H), 7.35–7.24 (m, 5 H), 5.21 (dd, $J = 6.0, 3.4$ Hz, 1 H), 4.69 and 4.48 (AB dd, $J = 11.0$ Hz, 1 H), 4.15 (ddd, $J = 8.3, 3.6, 3.4$ Hz, 1 H), 3.92 (qd, $J = 6.4, 6.0$ Hz, 1 H), 2.77 (ddd, $J = 16.9, 8.3, 2.4$ Hz, 1 H), 2.54 (ddd, $J = 16.9, 3.6, 1.6$ Hz, 1 H), 2.09 (s, 3 H), 1.14 (d, $J = 6.4$ Hz, 3 H), 0.88 (s, 9 H), 0.071 (s, 3 H), 0.054 (s, 3 H); IR (neat) 2960, 2860,

2710, 1745, 1725 cm^{-1} ; HRMS for $C_{15}H_{19}O_5$ ($M + TBDMS$), calcd 279.1232, found 279.1242.

A 0 °C solution of aldehyde 22 (173 mg, 0.44 mmol) in CH_3CN (2 mL) was treated with 10 drops of 48% HF in H_2O (ca. 0.89 mmol). The mixture was stirred at 0 °C for 3 h and then was diluted with water (20 mL) and extracted with $CHCl_3$ (2 × 25 mL). The organic extracts were dried ($MgSO_4$), filtered, and concentrated in vacuo. Purification of the crude product by flash column chromatography on silica gel (35% EtOAc/hexanes) gave 110 mg (90%) of viscous 23 as an anomeric mixture (ca. 2:1 ratio in favor of α -OH anomer), 7.4 mg (6%) of α,α -dimer 25, and 5 mg (4%) of impure α,β -dimer 26.

Data for 23: R_f (0.35 in 35% EtOAc-hexane); 1H NMR (500 MHz, $CDCl_3$); α -OH anomer δ 7.38–7.26 (m, 5 H), 5.44 (br s, 1 H), 5.35 (d, $J = 2.7$ Hz, 1 H), 4.70 and 4.44 (AB dd, $J = 11.4$ Hz, 1 H), 4.22 (q, $J = 6.6$ Hz, 1 H), 3.99 (ddd, $J = 11.8, 5.3, 2.7$ Hz, 1 H), 2.68 (dd, $J = 2.4, 2.4$ Hz, 1 H for OH), 2.17 (s, 3 H), 2.00 (dddd, $J = 13.0, 11.8, 3.4, 2.4$ Hz, 1 H), 1.94 (dd, $J = 13.0, 5.3$ Hz, 1 H), 1.16 (d, $J = 6.6$ Hz, 3 H); β -OH anomer δ 7.38–7.26 (m, 5 H), 5.25 (d, $J = 2.9$ Hz, 1 H), 4.76 (ddd, $J = 9.4, 7.1, 2.2$ Hz, 1 H), 4.69 and 4.46 (AB dd, $J = 11.8$ Hz, 1 H), 3.61 (q, $J = 6.5$ Hz, 1 H), 3.58 (ddd, $J = 12.2, 4.6, 2.9$ Hz, 1 H), 3.30 (d, $J = 7.1$ Hz, 1 H for OH), 2.18 (s, 3 H), 2.10 (ddd, $J = 12.6, 4.6, 2.2$ Hz, 1 H), 1.80 (ddd, $J = 12.6, 12.2, 9.4$ Hz, 1 H), 1.23 (d, $J = 6.5$ Hz, 3 H); IR (neat, for the mixture) 3400, 1725 cm^{-1} ; HRMS (CI) for $C_{15}H_{21}O_5$ ($M^+ + 1$) for the mixture, calcd 281.1389, found 281.1409. Anal. Calcd for $C_{15}H_{20}O_5$: C, 64.27; H, 7.19. Found: C, 63.99; H, 7.03.

Data for α,α -dimer 25: mp 114–116 °C; $[\alpha]_D^{25} +218^\circ$ ($c = 1.54$, $CHCl_3$); R_f 0.23 (30% EtOAc-hexane); 1H NMR (500 MHz, $CDCl_3$) δ 7.38–7.27 (m, 5 H), 5.35 (br d, $J = 3.7$ Hz, 1 H), 5.24 (d, $J = 3.0$ Hz, 1 H), 4.72 and 4.43 (AB dd, $J = 11.3$ Hz, 2 H), 3.92 (q, $J = 6.5$ Hz, 1 H), 3.88 (ddd, $J = 12.1, 4.8, 3.0$ Hz, 1 H), 2.17 (s, 3 H), 2.07 (ddd, $J = 12.9, 12.1, 3.7$ Hz, 1 H), 1.79 (ddd, $J = 12.9, 4.8, 1.1$ Hz, 1 H), 1.16 (d, $J = 6.5$ Hz, 3 H); IR (neat) 1735 cm^{-1} ; HRMS for $C_{21}H_{27}O_7$ ($M - C_7H_7 - HOAc$), calcd 391.1756, found 391.2844.

Partial data for α,β -dimer 26: R_f 0.32 (30% EtOAc-hexane); 1H NMR (500 MHz, $CDCl_3$) δ 7.39–7.27 (m, 10 H), 5.34 (br d, $J = 3.0$ Hz, 1 H), 5.24 (d, $J = 3.3$ Hz, 1 H), 5.18 (br s, 1 H), 4.69 and 4.42 (A'B' dd, $J = 11.3$ Hz, 2 H), 4.68 and 4.45 (A''B'' dd, $J = 12.0$ Hz, 1 H), 4.60 (dd, $J = 9.8, 2.4$ Hz, 1 H), 4.30 (q, $J = 6.6$ Hz, 1 H), 4.01 (ddd, $J = 10.9, 6.3, 3.0$ Hz, 1 H), 3.59 (qd, $J = 6.4, 1.0$ Hz, 1 H), 3.55 (ddd, $J = 12.1, 4.9, 3.3$ Hz, 1 H), 2.18 (s, 3 H), 2.16 (s, 3 H), 2.02–1.85 (m, 4 H), 1.20 (d, $J = 6.4$ Hz, 3 H), 1.11 (d, $J = 6.6$ Hz, 3 H); IR (neat) 1735 cm^{-1} .

tert-Butyltrimethylsilyl 4-O-Acetyl-3-O-benzyl-2,6-dideoxy-D-lyxo-pyranoside (24). To a 0 °C solution of lactol 23 (283 mg, 1.01 mmol) in CH_2Cl_2 (20 mL) were added 2,4-lutidine (0.35 mL, 3.03 mmol) and TBDMS-OTf (0.28 mL, 1.21 mmol). The mixture was stirred for 50 min at 0 °C and then was diluted with CH_2Cl_2 (150 mL) and H_2O (30 mL). The aqueous layer was extracted with CH_2Cl_2 (2 × 30 mL). The combined organic layers were washed with brine (20 mL) and dried over $MgSO_4$. Filtration, concentration of the filtrate in vacuo, and chromatography of the residue on silica gel (10% EtOAc/hexanes) gave 354 mg of 24 (89%) as a mixture of anomers. Samples of the two isomers were separated by careful multiple elution preparative TLC for spectroscopic characterization, but on a routine basis this mixture was used in the next transformation without separation.

Data for α -pyranoside 24a: R_f 0.57 (10% EtOAc/hexane, three elutions); $[\alpha]_D^{25} +116^\circ$ ($c = 0.74$, $CHCl_3$); 1H NMR (500 MHz, $CDCl_3$) δ 7.37–7.24 (m, 5 H), 5.35 (dd, $J = 3.0, 1.3$ Hz, 1 H), 5.33 (dd, $J = 3.1, 1.3$ Hz, 1 H), 4.71 and 4.44 (AB dd, $J = 11.4$ Hz, 1 H), 4.09 (qd, $J = 6.6, 1.3$ Hz, 1 H), 3.96 (ddd, $J = 12.3, 4.8, 3.1$ Hz, 1 H), 2.16 (s, 3 H), 2.00 (ddd, $J = 12.3, 12.3, 3.0$ Hz, 1 H), 1.79 (ddd, $J = 12.3, 4.8, 1.3$ Hz, 1 H), 1.13 (d, $J = 6.6$ Hz, 3 H), 0.87 (s, 9 H), 0.083 (s, 3 H), 0.076 (s, 3 H); IR (neat) 2950, 1740 cm^{-1} ; HRMS for $C_{17}H_{25}O_5Si$ ($M^+ - C_4H_9$), calcd 337.1471, found 337.1466. Anal. Calcd for $C_{21}H_{34}O_5Si$: C, 63.92; H, 8.68. Found: C, 64.30; H, 8.46.

Data for β -pyranoside 24b: R_f 0.66 (10% EtOAc/hexane three elutions); $[\alpha]_D^{25} +45.9^\circ$ ($c = 1.03$, $CHCl_3$); 1H NMR (500 MHz, $CDCl_3$) δ 7.38–7.27 (m, 5 H), 5.22 (ddd, $J = 3.1, 1.2, 0.9$ Hz, 1 H), 4.73 (dd, $J = 9.4, 3.0$ Hz, 1 H), 4.68 and 4.44 (AB dd, $J = 11.8$ Hz, 1 H), 3.57–3.52 (m, 2 H), 2.17 (s, 3 H), 1.95 (dddd, $J = 12.3,$

4.7, 3.0, 0.9 Hz, 1 H), 1.86 (ddd, $J = 12.3, 12.3, 9.4$ Hz, 1 H), 1.21 (d, $J = 6.3$ Hz, 3 H), 0.90 (s, 9 H), 0.13 (s, 3 H), 0.10 (s, 3 H); IR (neat) 2950, 2850, 1735 cm^{-1} ; HRMS for $\text{C}_{17}\text{H}_{25}\text{O}_5\text{Si}$ ($M^+ - \text{C}_4\text{H}_9$), calcd 337.1471, found 337.1467.

tert-Butyldimethylsilyl 4-O-Acetyl-2,6-dideoxy-D-lyxo-pyranoside (27). A solution of benzyl ether 24 (304 mg, 0.77 mmol; an anomeric mixture) in EtOH (3 mL) containing ca. 20 mg of 10% Pd-C was stirred under H_2 (1 atm) for 2 days at 23 °C. The solution was filtered and concentrated in vacuo. Chromatography of the residue on silica gel (25% EtOAc/hexanes) gave 62 mg of recovered 24 (20%) and 179 mg of 27 (76% yield; 93% based on consumed 24).

The hydrogenolysis reaction has been performed by using purified samples of the α - and β -anomers of 24, or on mixtures of them as described here, and similar results were obtained in all cases. The two anomers can be partially separated by silica gel chromatography, but mixtures were routinely used in the coupling reaction with thioglycoside 18.

Data for α -pyranoside 27 α : R_f 0.36 (30% EtOAc/hexane); mp 83–85 °C (solidified on standing); $[\alpha]_D^{25} +99.1^\circ$ ($c = 1.00$, CHCl_3); $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 5.35 (br s, 1 H), 5.04 (d, $J = 2.7$ Hz, 1 H), 4.32–4.26 (m, 1 H), 4.13 (q, $J = 6.6$ Hz, 1 H), 2.19 (d, $J = 4.9$ Hz, 1 H for OH), 2.17 (s, 3 H), 1.89–1.80 (m, 2 H), 1.13 (d, $J = 6.6$ Hz, 3 H), 0.89 (s, 9 H), 0.095 (s, 3 H), 0.083 (s, 3 H); IR (neat) 3440, 1730 cm^{-1} ; HRMS for $\text{C}_{14}\text{H}_{27}\text{O}_5\text{Si}$ ($M^+ - 1$), calcd 303.1628, found 303.1632. Anal. Calcd for $\text{C}_{14}\text{H}_{28}\text{O}_5\text{Si}$: C, 55.23; H, 9.27. Found: C, 55.60; H, 8.89.

Data for β -pyranoside 27 β : R_f 0.35 (30% EtOAc/hexane); $[\alpha]_D^{25} -3.0^\circ$ ($c = 0.97$, CHCl_3); $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 4.94 (br d, $J = 3.3$ Hz, 1 H), 4.74 (dd, $J = 9.4, 2.1$ Hz, 1 H), 3.93–3.87 (m, 1 H), 3.59 (qd, $J = 6.5, 1.1$ Hz, 1 H), 2.17 (s, 3 H), 2.01 (d, $J = 5.2$ Hz, 1 H for OH), 1.97 (dddd, $J = 12.2, 4.7, 2.1, 0.9$ Hz, 1 H), 1.72 (d, $J = 12.5, 12.2, 9.4$ Hz, 1 H), 1.21 (d, $J = 6.5$ Hz, 3 H), 0.90 (s, 9 H), 0.13 (s, 3 H), 0.11 (s, 3 H); IR (neat) 3450 (br), 1730 cm^{-1} ; HRMS for $\text{C}_{14}\text{H}_{27}\text{O}_5\text{Si}$ ($M^+ - 1$), calcd 303.1628, found 303.1643. Anal. Calcd for $\text{C}_{14}\text{H}_{28}\text{O}_5\text{Si}$: C, 55.23; H, 9.27. Found: C, 55.05; H, 9.27.

Synthesis of Thioglycoside 18 from 23. To a solution of lactol 23 (270 mg, 0.96 mmol) in CH_2Cl_2 (6 mL) was added TMS-SPh (912 mL, 4.8 mmol), Bu_4NI (0.43 g, 1.16 mmol) and ZnI_2 (0.92 g, 2.89 mmol). The mixture was stirred at 23 °C for 3 h, diluted with CH_2Cl_2 (50 mL), and poured into H_2O (50 mL). The aqueous layer was extracted with CH_2Cl_2 (2 \times 50 mL). The combined organic layers were dried (MgSO_4), filtered, and concentrated in vacuo. Chromatography of the residue on silica gel (80% CH_2Cl_2 /hexanes, followed by 20% EtOAc/hexanes) gave 333 mg (93%) of thioglycoside 28 as ca. 3:1 mixture in favor of the α -anomer. The anomers were separated by preparative TLC for spectroscopic characterization, but mixtures were routinely used in the coupling reaction.

Data for α -D-pyranoside 28 α : R_f 0.37 (15% EtOAc/hexanes); $[\alpha]_D^{24} +310^\circ$ ($c = 1.98$, CHCl_3); $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.46–7.40 (m, 2 H), 7.38–7.24 (m, 8 H), 5.74 (dd, $J = 5.7, 0.9$ Hz, 1 H), 5.39 (dd, $J = 3.0, 1.0$ Hz, 1 H), 4.72 and 4.48 (AB dd, $J = 11.4$ Hz, 2 H), 4.46 (q, $J = 6.5$ Hz, 1 H), 3.92 (ddd, $J = 12.4, 4.7, 3.0$ Hz, 1 H), 2.42 (ddd, $J = 13.4, 12.4, 5.7$ Hz, 1 H), 2.17 (s, 3 H), 2.11 (dddd, $J = 13.4, 4.7, 1.0, 0.9$ Hz, 1 H), 1.18 (d, $J = 6.5$ Hz, 3 H); IR (neat) 1735, 1560 cm^{-1} ; MS (CI) m/z 373 (M^+).

Data for the β -D-pyranoside 28 β : R_f 0.29 (15% EtOAc/hexanes); $[\alpha]_D^{24} +12.8^\circ$ ($c = 0.57$, CHCl_3); $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.53–7.49 (m, 2 H), 7.37–7.24 (m, 8 H), 5.28 (br d, $J = 3.0$ Hz, 1 H), 4.75 (dd, $J = 11.2, 2.9$ Hz, 1 H), 4.67 and 4.46 (AB dd, $J = 11.9$ Hz, 2 H), 3.62 (qd, $J = 6.4, 1.1$ Hz, 1 H), 3.60 (ddd, $J = 10.9, 5.6, 3.0$ Hz, 1 H), 2.17 (s, 3 H), 2.12–2.02 (m, 2 H), 1.26 (d, $J = 6.4$ Hz, 3 H); IR (neat) 1735, 1560 cm^{-1} ; HRMS for $\text{C}_{15}\text{H}_{19}\text{O}_4$ ($M^+ - \text{SPh}$), calcd 263.1283, found 263.1272.

To a solution of 28 (328 mg, 0.88 mmol; a mixture of anomers) in Me_2SO (3 mL) was added powdered KOH (0.28 g, 5.0 mmol). The mixture (yellow) was stirred at 23 °C for 1 h. Methyl iodide (0.33 mL, 5.3 mmol) was then added and the solution again turned clear. The reaction was stirred overnight, then was diluted with H_2O (50 mL), and extracted with CH_2Cl_2 (3 \times 50 mL). The organic extracts were washed with H_2O (2 \times 50 mL), dried (MgSO_4) filtered, and concentrated in vacuo. Chromatography of the residue on silica gel (13% EtOAc/hexanes) provided 255 mg (84%) of the thioglycoside 18 (anomeric mixture) as a yellow

liquid. This material was indistinguishable from samples of 18 prepared from 17 (see supplementary material). The following data are for a 25:75 mixture of α - and β -anomers: $R_f = 0.55$ in 20% EtOAc/hexanes; $[\alpha]_D^{25} +157^\circ$ ($c = 0.40$, CHCl_3); $^1\text{H NMR}$ (500 MHz, CDCl_3) α -D-pyranoside δ 7.52–7.20 (m, 10 H), 5.73 (dd, $J = 5.5, 1.1$ Hz, 1 H), 4.65 and 4.64 (AB dd, $J = 11.9$ Hz, 2 H), 4.27 (q, $J = 6.5$ Hz, 1 H), 3.86 (ddd, $J = 12.2, 4.5, 2.5$ Hz, 1 H), 3.63 (s, 3 H), 3.40 (dd, $J = 2.5, 1.3$ Hz, 1 H), 2.49 (ddd, $J = 13.1, 12.2, 5.5$ Hz, 1 H), 2.11 (dddd, $J = 13.1, 4.5, 1.3, 1.1$ Hz, 1 H), 1.26 (d, $J = 6.5$ Hz, 3 H); β -D-pyranoside δ 7.52–7.20 (m, 10 H), 4.70–4.67 (m, 1 H), 4.64 and 4.60 (AB dd, $J = 12.2$ Hz, 2 H), 3.62 (s, 3 H), 3.56–3.52 (m, 1 H), 3.44 (qd, $J = 6.4, 0.8$ Hz, 1 H), 3.30 (br d, $J = 2.5$ Hz, 1 H), 2.18–2.12 (m, 2 H), 1.33 (d, $J = 6.4$ Hz, 3 H); IR (neat for the mixture) 3020, 2920, 1560, 1475, 1365, 1200, 1080, 720 cm^{-1} ; MS m/z 344 (M^+). Anal. Calcd for $\text{C}_{20}\text{H}_{24}\text{O}_3\text{S}$ (for the mixture): C, 69.74; H, 7.02. Found: C, 69.48; H, 6.74.

tert-Butyldimethylsilyl 4-O-Acetyl-3-(3-O-benzyl-2,6-dideoxy-4-O-methyl- α -D-lyxo-pyranosyl)-2,6-dideoxy-D-lyxo-pyranoside (4b). A mixture of thioglycoside 18 (16 mg, 0.046 mmol), purified alcohol 27 α (15 mg, 0.049 mmol), and 4-Å molecular sieves (ca. 20 mg) in CH_3CN -THF (3 mL, 2:1) was cooled to –42 °C. Recrystallized NBS (12 mg, 0.070 mmol) was then added. The mixture was stirred at –42 °C for 50 min and then added. The mixture was stirred at –42 °C for 50 min and then was diluted with saturated aqueous NaHCO_3 solution (13 mL) and extracted with EtOAc (3 \times 20 mL). The organic extracts were washed with saturated NaHCO_3 solution (2 \times 20 mL) and brine (2 \times 20 mL), dried (MgSO_4), filtered, and concentrated in vacuo. Purification of the residue by preparative TLC (0.5-mm plate 15% EtOAc/hexanes, four elutions) gave 23 mg of α,α -disaccharide 4b (92%).

This reaction has been run by using purified samples of 27 α , 27 β , or mixtures of them, and similar results were obtained in all cases. In one experiment with 27 β , a small amount (4%, but still contaminated with some other minor impurities) of an isomer that was identified as the β,β -isomer of 4b. Stereochemical assignments and the following ^1H and ^{13}C NMR assignments are based on 2D NMR ^1H - ^1H correlation experiments, ^1H - ^{13}C correlation experiments, and ^1H decoupling experiments.

Data for α,α -isomer of 4b: R_f 0.60 (30% EtOAc-hexane); $[\alpha]_D^{27} +156^\circ$ ($c = 0.96$, CHCl_3); $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.37–7.25 (m, 5 H), 5.34 (br s, 1 H, H_1), 5.15 (br s, 2 H, H_4 , H_1), 4.59 and 4.58 (AB, $J = 11.9$ Hz, 2 H), 4.24 (ddd, $J = 12.2, 4.7, 3.0$ Hz, 1 H, H_3), 4.10 (q, $J = 6.6$ Hz, 1 H, H_5), 3.80 (ddd, $J = 12.0, 4.6, 2.6$ Hz, 1 H, H_3), 3.74 (q, $J = 6.5$ Hz, 1 H, H_5), 3.60 (s, 3 H), 3.33 (br s, 1 H, H_4), 2.12 (s, 3 H), 2.04 (ddd, $J = 12.8, 12.0, 3.8$ Hz, 1 H, H_{2ax}), 1.95 (ddd, $J = 12.3, 12.2, 3.1$ Hz, 1 H, H_{2ax}), 1.77 (dd, $J = 12.8, 4.6$ Hz, 1 H, H_{2eq}), 1.60 (dd, $J = 12.3, 4.7$ Hz, 1 H, H_{2eq}), 1.22 (d, $J = 6.5$ Hz, 3 H, H_6), 1.10 (d, $J = 6.6$ Hz, 3 H, H_6), 0.89 (s, 9 H), 0.094 (s, 3 H), 0.085 (s, 3 H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 170.8, 138.7, 128.3, 127.4, 127.2, 94.6 (C_1), 92.5 (C_1), 78.6 (C_4), 74.7 (C_3), 70.1, 69.0 (C_4), 66.8 (C_5), 66.6 (C_3), 64.7 (C_5), 61.4 (C_7), 33.8 (C_2), 30.1 (C_2), 25.6, 20.8, 17.9, 16.8 (C_6), 16.7 (C_6), –4.6, –5.9; IR (neat) 2950, 2860, 1740 cm^{-1} ; HRMS for $\text{C}_{24}\text{H}_{37}\text{O}_8\text{Si}$ ($M^+ - \text{C}_4\text{H}_9$), calcd 481.2258, found 481.2243.

Data for α,β -isomer of 4b: R_f 0.43 (30% EtOAc-hexane); $[\alpha]_D^{24} +86.6^\circ$ ($c = 1.55$, CHCl_3); $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.37–7.26 (m, 5 H), 5.09 (br d, $J = 3.7$ Hz, 1 H, H_1), 5.03 (dd, $J = 3.2, 1.1$ Hz, 1 H, H_4), 4.75 (ddd, $J = 8.1, 3.5, 1.0$ {from decoupling} Hz, 1 H, H_1), 4.59 and 4.57 (AB dd, $J = 12.0$ Hz, 2 H), 3.83–3.76 (m, 3 H, H_3 , H_3 , H_5), 3.61 (s, 3 H), 3.55 (qd, $J = 6.4, 1.1$ Hz, 1 H, H_5), 3.32 (br s, 1 H, H_4), 2.10 (s, 3 H), 2.03 (ddd, $J = 12.4, 12.4, 3.7$ Hz, 1 H, H_{2ax}), 1.78 (ddd, $J = 12.3, 12.3, 8.1$ Hz, 1 H, H_{2ax}), 1.77–1.74 (m, 2 H, H_{2eq} , H_{2eq}), 1.24 (d, $J = 6.6$ Hz, 3 H, H_6), 1.18 (d, $J = 6.4$ Hz, 3 H, H_6), 0.91 (s, 9 H), 0.13 (s, 3 H), 0.11 (s, 3 H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 170.9, 138.6, 128.3, 127.5, 127.2, 95.7 (C_1), 95.0 (C_1), 78.6 (C_4), 74.5, 70.7, 70.1, 69.2 (C_5), 67.7 (C_4), 67.0, 61.4, 36.1 (C_2), 30.1 (C_2), 25.8, 20.9, 18.1, 17.0 (C_6), 16.7 (C_6), –4.1, –5.0; IR (CHCl_3) 3020, 2915, 2850, 1730 cm^{-1} ; HRMS for $\text{C}_{28}\text{H}_{45}\text{O}_8\text{Si}$ ($M^+ - 1$), calcd 537.2884, found 537.2888.

Partial data for β,β -isomer of 4b: R_f 0.27 (30% EtOAc/hexane); $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.37–7.25 (m, 5 H), 5.04 (d, $J = 3.3$ Hz, 1 H), 4.71 (dd, $J = 9.5, 2.2$ Hz, 1 H, H_1), 4.60 and 4.57 (AB dd, $J = 12.2$ Hz, 1 H), 4.46 (dd, $J = 8.8, 3.1$ Hz, 1 H, H_1), 3.95 (ddd, $J = 12.3, 4.5, 3.3$ Hz, 1 H), 3.59 (s, 3 H), 3.52 (q, $J = 6.4$ Hz, 1 H), 3.47–3.43 (m, 1 H), 3.34 (q, $J = 6.5$ Hz, 1 H), 3.21 (br s, 1 H), 2.09 (s, 3 H), 1.97–1.92 (m, 1 H), 1.92–1.85 (m, 2 H),

1.74 (ddd, $J = 12.3, 12.3, 9.5$ Hz, 1 H), 1.26 (d, $J = 6.4$ Hz, 3 H), 1.13 (d, $J = 6.5$ Hz, 3 H), 0.88 (s, 9 H), 0.11 (s, 3 H), 0.086 (s, 3 H).

4-O-Acetyl-3-O-(3-O-benzyl-2,6-dideoxy-4-O-methyl- α -D-lyxo-pyranosyl)-2,6-dideoxy-D-lyxo-pyranose (4c). A solution of **4b** (40 mg, 0.075 mmol) and $\text{Et}_3\text{NH}^+\text{F}^-$ (8 equiv) in CH_3CN (2 mL) was heated to 70 °C for 2.5 h. Saturated NaHCO_3 solution (0.5 mL) was added and the mixture was stirred at 23 °C for 10 min. The solution was diluted with additional aqueous NaHCO_3 (10 mL) and was extracted with EtOAc (4×20 mL). The organic extracts were washed with saturated NaHCO_3 solution (20 mL) and brine (2×20 mL) and dried over MgSO_4 . After filtration and concentration of the filtrate in vacuo, the residue was purified with preparative TLC (0.5-mm plate, 50% EtOAc /hexanes, two elutions), giving 32 mg (100%) of disaccharide **4c** as ca. 2:1 mixture in favor of α -OH anomer.

This reaction has been run with the purified α,α - and α,β -anomers of **4b**, or on mixtures of them; the yields have always been quantitative. It was noticed that the β -TBDMS anomer of **4b** was desilylated faster when a mixture was used. The following ^1H and ^{13}C NMR assignments are based on 2D NMR ^1H - ^1H correlation, ^1H - ^{13}C correlation, and ^1H decoupling experiments.

Data for **4c**: R_f 0.20 (50% EtOAc /hexane); ^1H NMR (500 MHz, CDCl_3) data for α -OH anomer δ 7.40-7.25 (m, 5 H), 5.40 (d, $J = 3.5$ Hz [after D_2O exchange], 1 H, H_1), 5.14 (d, $J = 2.6$ Hz, 1 H, H_4), 5.12 (d, $J = 3.8$ Hz, 1 H, H_1), 4.56 (s, 2 H), 4.23-4.17 (m, 2 H, H_3, H_5), 3.80-3.75 (m, 1 H, H_3 , overlapping with H_3 and H_5 of the β -anomer), 3.76 (q, $J = 6.5$ Hz, 1 H, H_5), 3.58 (s, 3 H), 3.31 (br s, 1 H, H_4), 2.85 (br s, 1 H, OH), 2.10 (s, 3 H), 2.02 (ddd, $J = 12.5, 12.5, 3.8$ Hz, 1 H, H_{2ax}), 1.89 (ddd, $J = 12.6, 12.6, 3.5$ Hz, 1 H, H_{2ax}), 1.73 (br dd, $J = 12.6, 4.9$ Hz, 2 H, $\text{H}_{2eq}, \text{H}_{2eq}$, almost superimposed), 1.22 (d, $J = 6.5$ Hz, 3 H, H_6), 1.10 (d, $J = 6.5$ Hz, 3 H, H_6); data for β -OH anomer δ 7.40-7.25 (m 5 H), 5.08 (d, $J = 3.6$ Hz, 1 H, H_1), 5.05 (d, $J = 3.2$ Hz, 1 H, H_4), 4.78 (ddd, $J = 9.6, 6.6, 1.9$ Hz, 1 H, H_1), 4.56 (s, 2 H), 4.25-4.17 (m, 1 H, H_3),

3.38 (ddd, $J = 12.3, 4.8, 3.2$ Hz, 1 H, H_3), 3.80-3.73 (m, 2 H, H_3, H_5), 3.58 (s, 3 H), 3.54 (d, $J = 6.6$ Hz, 1 H, OH), 3.31 (br s, 1 H, H_4), 2.09 (s, 3 H), 2.05-1.68 (m, 4 H, $\text{H}_{2ax}, \text{H}_{2ax}, \text{H}_{2eq},$ and H_{2eq}), 1.22 (d, $J = 6.5$ Hz, 3 H, H_6), 1.18 (d, $J = 6.4$ Hz, 3 H, H_6); ^{13}C NMR (125 MHz, CDCl_3) (for the mixture) δ 170.75 (carbonyl, for both anomers), 138.59, 138.49, 128.34, 128.32, 127.50, 127.45, 127.23, 127.21, 95.62, 94.99, 94.36, 92.37, 78.64, 78.54, 74.57, 74.40, 70.34, 70.11, 70.08, 69.42, 68.89, 67.50, 67.12, 66.98, 66.70, 64.72, 61.38, 61.36, 34.89, 31.42, 30.17, 30.06, 20.82, 20.77, 16.97 (C_6 , for both anomers), 16.78, 16.66; IR (neat) 3420, 3090, 3070, 2860, 2815, 1740 cm^{-1} ; HRMS for $\text{C}_{22}\text{H}_{30}\text{O}_7$ ($\text{M}^+ - \text{H}_2\text{O}$), calcd 406.1991, found 406.1974. Anal. Calcd for $\text{C}_{22}\text{H}_{32}\text{O}_8$: C, 62.25; H, 7.60. Found: C, 61.96; H, 7.64.

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Registry No. 1, 6988-58-5; 2, 7059-24-7; **4a**, 106023-24-9; α,α -**4b**, 131013-38-2; α,β -**4b**, 131013-40-6; β,β -**4b**, 131013-41-7; α -**4c**, 131041-37-7; β -**4c**, 131013-42-8; 8, 99603-55-1; 9, 127851-95-0; 10, 106062-13-9; 10 benzyl ether, 106062-14-0; 11, 106062-15-1; 11 TBDMS derivative, 131013-39-3; α -12, 107908-97-4; β -12, 131013-43-9; α -13, 131013-31-5; β -13, 131013-44-0; α -14, 4833-12-9; β -14, 106023-19-2; α -15, 106023-20-5; β -15, 106023-21-6; 16, 4092-40-4; 17, 3868-01-7; α -18, 106023-22-7; β -18, 106023-23-8; 19, 75810-18-3; 21, 131013-32-6; 22, 131013-33-7; α -23, 131013-34-8; β -23, 131013-45-1; α -24, 131013-35-9; β -24, 131013-46-2; 25, 131013-36-0; 26, 131100-33-9; α -27, 131041-38-8; β -27, 131013-47-3; α -28, 131013-37-1; β -28, 131013-48-4; 29, 85273-19-4; 30, 131100-34-0; 31, 131100-35-1.

Supplementary Material Available: Experimental procedures for the syntheses of **14a**-**19** and ^1H NMR spectra of **24** (α,β mixture), **25**, **26**, α,α -**4b**, β,α -**4b**, and β,β -**4b** (10 pages). Ordering information is given on any current masthead page.

Notes

Mechanistic Details for SET-Promoted Photoadditions of Amines to Conjugated Enones Arising from Studies of Aniline-Cyclohexenone Photoreactions

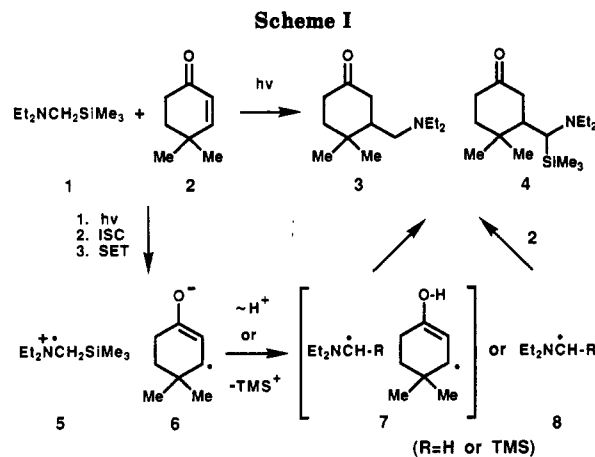
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Introduction

Previous studies exploring photoaddition reactions of α -silyl amines and α,β -unsaturated ketones have led to a number of interesting proposals concerning the mechanistic features of these single electron transfer (SET) promoted processes^{1,2} and have suggested potential synthetic applications.³ In mechanistic studies of the N -[(trimethylsilyl)methyl]- N,N -diethylamine (**1**) addition to 4,4-dimethylcyclohex-2-en-1-one (**2**), we observed that the relative yields of the non-TMS (**3**) and TMS adducts (**4**)



were dependent upon the nature of the solvent, metal cation additives, and amine concentration. These results were interpreted in terms of a mechanism (Scheme I) in which the relative rates of intermediate amine cation radical **5** deprotonation and desilylation are governed by the basicity of the enone anion radical **6**, which itself is controlled by hydrogen bonding and metal cation coordination. Furthermore, we suggested that in cases where the enone anion radical is rendered nonbasic by hydrogen bonding in polar protic solvents, amine cation radical **5**

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